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1Fabrication of cellulose carbamate hydrogel-dressing with rarasaponin surfactant for enhancing adsorption of silver nanoparticles and antibacterial activity

Vania Bundjajaa,

1Shella Permatasari Santosoc,**a,***, **Artik Elisa** Angkawijayab,**, **Maria**

Yulianac, Felycia Edi Soetaredjoc,a, Suryadi Ismadjic,a, Aning Ayucitrac, Chintya Gunartoa, Yi-Hsu Jua,b,d, Ming-Hua Hoa a Chemical Engineering Department, National Taiwan University of Science and Technology, #43, Sec. 4, Keelung Rd., Taipei 10607, Taiwan b Graduate Institute of Applied Science and Technology, National Taiwan University of Science and Technology, #43, Sec. 4, Keelung Rd., Taipei 10607, Taiwan c Department of Chemical Engineering, Widya Mandala Surabaya Catholic University, #37 Kalijudan Rd., Surabaya 60114, East Java, Indonesia d

5Taiwan Building Technology Center, National Taiwan University of Science and Technology, #43, Sec. 4, Keelung Rd., Taipei 10607, Taiwan

ARTICLE INFO ABSTRACT Keywords: Cellulose carbamate Silver nanoparticles Wound dressing Antibacterial dressing Bacterial contamination on external wounds is known to be a factor that prevents wound healing and triggers tissue damage. Hydrogel-dressings with antibacterial activity is a useful medical device to avoid this contamination, wherein the antibacterial activity can be provided via incorporation of silver nanoparticles (AgNPs). Contrary to the conventional two-step preparation of an AgNPs-loaded hydrogel (AgNPs@hydrogel), this work aims to establish a new and facile synthesis method employing the adsorption principle. Once AgNO₃ adsorbed into active sites of the hydrogels, in situ reductions using NaBH₄ was employed to produce AgNPs@hydrogel. The effect of surfactant addition on the AgNO₃ loading and the antibacterial activity of the resulting hydrogel dressing was investigated. The outcome of this work indicates that the addition of rarasaponin not only can increase the loading of AgNPs on cellulose carbamate hydrogel (CCH) but also significantly enhance the antibacterial activity of the resulted hydrogel-dressing. Superior to the other studied surfactant, the loading capacity (LC) of AgNPs is found to be 10.15, 9.94, and 7.53 mg/g for CCH modified with rarasaponin, CTAB, and Tween80, respectively. These findings conclude that the addition of surfactant, especially rarasaponin, can effectively improve the loading of AgNPs onto hydrogel-dressing via adsorption and promote the antibacterial activity. Furthermore, the cytotoxic test shows that the hydrogel-dressings have good biocompatibility toward skin fibroblast cells. 1. Introduction The development of antibacterial dressing for the treatment of external wounds has received considerable interest in recent years. The urge for this development is particularly triggered by the emergence of diabetes-related diseases, such as foot ulceration [1]. The existence of antibacterial properties in the wound dressing is crucial to prevent further tissue damage due to bacterial contamination and infection; these properties can be established by grafting antibacterial agents. Broad-spectrum antibacterial agents are usually used in wound dressing; these agents have complex mechanisms in interacting with bacterial cells, thereby avert the bacteria to develop resistance. Silver nanoparticles (AgNPs) are known as a powerful

broad-spectrum anti- microbial agent which have a long-lasting antimicrobial effect against various strains of bacteria and drug-resistant pathogens [2–7]. For centuries, AgNPs have been used to generate antimicrobial activity of different wound healing materials or devices, such as wound dressing, antiseptics, and medical devices [8]. Numerous techniques have been established to synthesis AgNPs, such as chemical reduction, photochemical, electrochemical, irradiation, thermal, biological, and microemulsion [9–18]. Among them, the chemical reduction is the most frequently adopted method owing to its *

Correspondence to: S. P. Santoso, Department of

4Chemical Engineering, Widya Mandala Surabaya Catholic University, #37 Kalijudan Rd., Surabaya 60114, East Java, Indonesia. Chemical Engineering Department, National Taiwan University of Science and Technology, #43, Sec. 4, Keelung Rd., Taipei 10607, Taiwan

. ** Corresponding author. E-mail addresses: sheila_p5@yahoo.com (S.P. Santoso),

14artikelisa@mail.ntust.edu.tw (A.E. Angkawijaya

). <https://doi.org/10.1016/j.msec.2020.111542> Received 15 April 2020; Received in revised form 7 September 2020; Accepted 19 September 2020 Available online 23 September 2020 0928-4931/ © 2020 Elsevier B.V. All rights reserved. concise and straightforward approach [19]. The tendency of AgNPs to form aggregates is a persistent problem in its synthesis, where the formation of aggregates can diminish the antibacterial efficiency of AgNPs [20]. Integration of AgNPs into supporting matrixes or polymeric material (like clay minerals, alginate-based hydrogel, chitosan, cellulose, collagen, PVA-PVP, and HEMA-PEGDA) is reportedly preventing the formation of the aggregates [21–31]. Hydrogels have gained much interest among these supporting materials, owing to their superior medical biocompatibility and immense water retention ability. Furthermore, hydrogels also can be directly applied as wound dressing materials [32,33]. Various natural and synthetic polymeric materials are widely available for preparing hydrogels, such as cellulose. In addition to the natural availability of cellulose, the beneficial biological properties of cellulose also promote its potency as raw material for wound dressing hydrogels. The beneficial biological properties of cellulose are including non-toxic, biodegradable, and high similarity in mechanical properties to natural tissue [34]. The biocompatibility and antimicrobial activity of cellulose in the form of hydrogel have been proven [35,36]. Herein, cellulose is utilized as the raw material for preparing hydrogel and also as a template for AgNPs nano-formulations. This combination can be a promising approach to prevent AgNPs aggregation while producing antimicrobial dressing. The preparation of cellulose hydrogel involves two main steps, that is the dissolution of the cellulose chain and the formation of a three-dimensional network via cross-linking. The nature of cellulose, which is not soluble in water, is a challenge in preparing cellulose-based hydrogels. Various dissolution methods have been established to resolve this limitation, such as the viscose process, carbamation, lyocell process, and the use of the ionic-liquid system [37]. Carbamation of cellulose to produce cellulose derivatives (i.e., cellulose carbamate) is known as an environmental dissolution method of cellulose compared to another soluble cellulose derivative (i.e., cellulose xanthogenate) which the synthesis accompanied by the generation of hazardous by-products. Cellulose carbamate (CC) has better solubility in alkaline and organic solution compared to cellulose [38]. Furthermore, CC reported to possess bacteriostatic properties, which make it

13more resistant against microbial contamination and enzymatic-cleavage compare to cellulose

. Gan et al. also reported that CC-based hydrogel has an enhanced porosity compared to cellulose hydrogels [39]. Therefore, in this work, the carbamation process was chosen to dissolve the cellulose. A new-facile approach in the incorporation of AgNPs to hydrogel was introduced. That is, by employing adsorption techniques of Ag⁺ ions onto the surface active sites of the hydrogel, followed by in situ reductions of the attached Ag⁺ ions. This approach allows the binding of large numbers of AgNP to the surface of the hydrogel dressing, thereby enhancing the antibacterial activity against the target bacteria. Furthermore, this approach could be an improvement over the conventional AgNPs hydrogel-dressing—wherein, in the preparation, AgNPs are integrated into the hydrogel matrix. So, the effectiveness of AgNPs in the center of the hydrogel matrix is reduced due to the difficulty of reaching the target bacteria. The effect of surfactant addition in improving the AgNPs loading capacity of the hydrogel was assessed in this work, where this case is yet to be significantly studied. The addition of surfactants also can enhance antibacterial properties. As reported by Cuahtecntzi-Delint et al., the incorporation of non-ionic surfactants (Triton X-100, Polyvinyl Pyrrolidone, and Tween80) on CeO₂ nanoparticles can enhance the antibacterial activity of the material [40]. An in vivo study by Sibbald et al. showed that a polymeric membrane foam pad filled with surfactant could overcome bacterial damage and facilitate wound closure [41]. Additionally, surfactant supplementation may significantly improve the stability of AgNPs [42,43]. The effect of three surfactants on the loading of AgNPs to CC-based hydrogel was assessed in this work. The surfactants used are (1) the non-ionic Tween80, (2) cationic CTAB, and (3) rarasaponin, which had no formal charge, but were deacylated when dissolved in polar solvents to negatively charged surfactants [44]. So far, there are still no studies that examine the effect of adding surfactants, especially rarasaponin, to increase the loading of AgNPs in CC-hydrogel (AgNPs@CCH). This work aims to investigate the effect of surfactants (especially rarasaponin) in the loading capacity of AgNPs to CCH. Some physicochemical measurements were carried out to characterize the material. The evaluation of cytotoxicity was carried out quantitatively using MTT assay. Hereinafter, the antibacterial activity of AgNPs@CCH with and without surfactant modification was investigated against Escherichia

10coli (gram-negative bacteria) and Staphylococcus aureus (gram-positive bacteria)

).

122. Materials and methods 2.1. Materials Laboratory **filter paper** (Whatman #1) **was used as a source**

of cellulose fibers. The following chemicals were used in this experiment: urea (CO(NH₂)₂, 98%; EMD Millipore, Darmstadt, Germany), sodium hydroxide (NaOH, 96%; EMD Millipore, Darmstadt, Germany), epichlorohydrin (ECH, ≥99%; Sigma-Aldrich, Singapore), silver nitrate (AgNO₃; ≥99.8%; Spectrum Chemical, Gardena, California), and sodium borohydride (NaBH₄, 99%; Sigma-Aldrich, Steinheim, Germany). The following surfactants were used for modification: polyoxyethylene sorbitan monooleate (Tween80; non-ionic surfactant; EMD Millipore, Darmstadt, Germany) cetyltrimethylammonium bromide (CTAB, cationic surfactant; > 98%; Sigma-Aldrich, Darmstadt, Germany), and Sapindus rarak DC fruit extract (rarasaponin; anionic surfactant;

3Sapindus rarak DC fruit obtained from Malang, East Java, Indonesia

). The structural properties and critical micelle concentration (CMC) of surfactants were presented in Supplementary Material Table S1.

3 Rarasaponin was extracted from Sapindus rarak according to

the previously published work [44], with a slight modification that is ethanol was used as the solvent instead of water. All solutions were prepared using reverse osmosis water obtained from Kusatsu water purification equipment with 8.3 $\mu\text{S}/\text{cm}$ conductivity at 28.5 °C. 2.2. Carbamation of cellulose The filter paper was cut into small pieces and pulp by soaking 1 g of filter paper in 100 mL of 3% NaOH solution for 1 h, with constant stirring at

17 room temperature. The pulp was washed with reverse osmosis (RO) **water** until **the**

washing solution pH reduces to 6. The pulp was then

4 dried in a 60 °C oven overnight. **The** carbamation process **was**

carried out by soaking 1 g of dry pulp in 20 mL of urea solution at a ratio of cellulose to urea of 1:1, 1:3, and 1:5. The mixture was stirred vigorously and then heated in an autoclave at 160 °C. After 1 h, the carbamation reaction was quenched by immersing the autoclave in a water bath filled with ice water. The resulting cellulose carbamate (CC) was washed several times with RO water to remove excess urea. Then, CC is dried in the oven overnight and stored in a desiccator. The resulting CC is coded as CC1, CC3, and CC5 according to the cellulose to urea ratio. 2.3. Preparation of CC hydrogel The alkaline solution was prepared by combining 7 wt% NaOH and 12 wt% urea, and then precooled at -13 °C. Into 100 g of the precooled solution, 4 wt% of CC was added and stirred until a transparent-viscous solution formed. The viscous

13 solution was then centrifuged at 5000 rpm for 3 min to remove trapped **air** **bubbles**

. Then, 5% (w/v) of ECH was added dropwise and stirred for 20

6 min. The gel solution was cast **into a** multi-well plate **and**

allowed to stand for about 15 min to condense the gel. The consolidation of the gel was initiated by heating

7 at 60 °C in a water bath. The resulting **hydrogel**

was continuously washed in flowing RO water for 2 d. Fig. 1. Schematic diagram of the process of (a) cellulose carbamation, (b)

7 dissolution of cellulose carbamate **in NaOH/urea**

, and (c)

7 cross-linking of cellulose carbamate with ECH

. Fig. 2. Optical properties of CCH (a) digital photos that show the level of transparency, from left to right: MFP, CC1, CC3, and CC5, and (b) UV–Vis transmittance spectrum. Table 1 Effect of surfactant on pH_{pzc} of CCH. Modifying surfactant Surfactant conc. (mM) Sample code pH_{pzc} – Tween80 CTAB Rarasaponin – CCH 5 CCH-T5 10 CCH-T10 5 CCH-C5 10 CCH-C10 5 CCH-R5 10 CCH-R10 6.18 6.45 6.34 7.78 8.03 5.54 5.25 2.4. Modification of CC hydrogel using surfactants Rarasaponin, Tween80, and CTAB are used as surfactants to modify CC hydrogel (CCH). Briefly, 10 mL of surfactant at a concentration of 5 or 10 mM was prepared in distilled water. The freeze-dried hydrogel disk (10 mm diameter, 2 mm height, and dry weight ± 10 mg) was then soaked in the surfactant solution for 48 h at room temperature. The hydrogel was then rinsed with RO water and then dried using a freeze-dryer. The surfactant modified hydrogels were denoted as CCH- x, with x = T, C, or R according to the type of surfactant Tween80, CTAB, and rarasaponin, respectively. Fig. 3. SEM images AgNPs-loaded (

9a–b) CCH, (c–d) CCH-T, (e–f) CCH-C, and (g–h

) CCH-R. The insets in panels show the physical appearance of hydrogels after drying. (

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.) 2.5. AgNPs embedding into CCH The formation of AgNPs was carried out using chemical reduction techniques with NaBH₄ as its reducing agent [45]. Hydrogel discs (with or without surfactant modification)

17 were immersed in 10 mL of 10 mM AgNO₃ solution

. After 24 h immersion, the Ag⁺-containing hydrogels were transferred into 10 mL of 10 mM NaBH₄ solution and left for 2 h. The obtained AgNPs-loaded CCH was washed using RO water and subsequently dried using a freeze-dryer. The concentration of em- bedded Ag⁺ ions was measured using an atomic absorption spectro- photometer (AAS) against AgNO₃ in 0.5 M HNO₃ as the standard. 2.6. Characterization and analysis 2.6.1. Characterizations Transparency measurement of CCH was carried out by pouring the gel into a quartz cuvette with a 10 mm pathlength. Then the trans- mittance spectra were recorded using UV–Vis Spectrophotometer (Shimadzu UV-1700) at 850–250 nm wavelength with 400 nm/min scanning speed. The cross-sectional images and subsequent elemental diagram of the

1 samples were taken using a field-emission scanning electron microscope

(JEOL JSM-6390) with an attachment of energy dispersive spectroscopy (EDS) analyzer; images were taken at an ac- celerating

7 voltage of 20 kV. Before SEM imaging, samples were freeze- dried

and plunged into liquid nitrogen. The samples were then broken into small fractures; fractures were then coated with platinum by a fine auto coater (JEC-3000FC). The morphology of AgNPs in the hydrogels was

observed by the use of FEI Tecnai G2 TF20 Super TWIN. The TEM specimens were prepared by crushing the samples and dispersing in ethanol. Sample solutions were deposited on

15 **a carbon-coated copper grid and allowed to sit to evaporate the**

solvent prior to the imaging. The dimension of AgNPs from TEM micrographs was measured using ImageJ 1.52a software. Surface functional groups were characterized using Fourier transformed infrared (FTIR) spectra. The FTIR pattern was accumulated from 20 scans using a Shimadzu 8400S spectro- photometer equipped with a KBr beam splitter in 4000–400 cm^{-1} re- gion. The crystal diffraction pattern was recorded by

12 **Bruker D2 Phaser X-ray diffractometer with a monochromatic high-intensity Cu K α**

ra- diation

1 **operating at 30 kV and 10 mA. 2**

.6.2. Surface charge measurement (pHpzc) The pHpzc analysis was performed as described by Orfao et al. (2006), with minor modifications [46].

16 **A series of capped Erlenmeyer flask containing 10 mL NaCl solution**

(0.01 M) were prepared.

16 **Each flask was adjusted to a specific pH**

, at a pH range of 2 to 10, by the addition of HCl and NaOH. To each flask, a 30 mg sample was added. The sample was immersed for 48 h with constant shaking at 100 rpm and a constant temperature of 30 °C. The final pH of each solution was measured using a pH meter (Mettler-Toledo S20 SevenEasy). The pH_{final} versus pH_{initial} values were plotted, the corresponding intersection point of these two curves (at pH_{final} = pH_{initial}) was identified as pHpzc. 2.6.3. Measurement of loaded AgNPs The amount of Ag loading on CCH was measured using atomic absorption spectrophotometric (AAS) (Shimadzu AA-6200) equipped with an autosampler, silver hollow cathode lamp, and air-acetylene flame. The AgNPs-hydrogel discs were first dissolved in 5 mL of aqua regia (prepared by mixing HCl and HNO₃ solution at ratio 3:1 (v/v)). The Fig. 4. (a) X-ray diffraction (XRD) pattern of pristine filter paper, AgNPs@CCH, and surfactant modified AgNPs@CCH. (b) SAED pattern of AgNPs@CCH-R10. AgNPs loading capacity (LC) of the hydrogel is calculated using the following formula: $LC (\mu\text{g AgNP/mg dry hydrogel}) = C \times V / m$ (LC \times m) where C ($\mu\text{g/mL}$) is the amount of AgNPs in the solution, V (mL)

5 **is the volume of the solution, and m (mg) is the mass of**

dry hydrogel along with the loaded AgNP. The divider on the right needs to be subtracted by the loaded-AgNPs weight, leaving only dry hydrogel mass (without AgNPs). A goal-seek function in Microsoft Excel was

used to solve the LC mass balance. 2.6.4. Deformability measurement Penetrometer (Ashoka Enterprises) was employed to measure the deformability of swollen hydrogels at 30 °C. A needle with an attached load of 50 g with a square-shaped ends precisely adjusted to touch the surface of the cylindrical hydrogel (10 mm in diameter, 2 mm in height). Then, the needle was allowed to sink into the sample for 5 s. The values shown on the scale were recorded, and the deformability was expressed in mm/g s. 2.6.5. Cytotoxicity assay The cytotoxicity of hydrogel samples was evaluated in vitro toward the skin fibroblast, L929 cell, by 3-

194,5-dimethyldiazol-2-yl)-2,5-di- phenyltetrazolium bromide (MTT) assay.
Dulbecco's modified eagle's medium

(DMEM) was used as the culture media. Cells were seeded as a monolayer in culture flasks at 37 °C ± 1 °C, 90% ± 10% humidity, and 5.0% ± 1.0% CO₂/air. After 24 h of incubation, the medium was substituted with a serum-free medium containing prepared hydrogel extract (0.2 g hydrogel in 1 mL medium) according to ISO 10993-5 and then incubated for 24 h at 37 °C. Following, 5 mg/mL of MTT was added and incubated for a further 4 h. After removing the medium, DMSO was added and shaken for 30 min. The absorbance of the purple formazan crystals was recorded at 570 nm using an automatic micro- plate reader (ELx800; Bio-Tek Instruments, Winooski, VT, USA). 2.6.6. Antibacterial activity assay The disk diffusion method was applied to assay the antibacterial activity of hydrogel samples. The inhibitory effect of the samples was tested against two bacterial strains, that is

10Escherichia coli (E. coli, ATCC 25922; gram-negative) and Staphylococcus aureus (S. aureus, ATCC 25923; gram-positive

). Before the assay, bacterial suspension at a concentration of 10⁸ CFU/mL was prepared according to the 0.5 McFarland standard. The bacterial suspension was then inoculated onto Mueller-Hinton agar plates using a sterile cotton swab. The freeze-dried AgNP-hydrogel samples were pressed into discs with a diameter of 10 mm. The pressed hydrogel discs were placed on the surface of the agar plates aseptically. The inhibition zone was measured after 24 h incubation at 37 °C. Ampicillin (10 µg) and AgNP-free hydrogels were used as a positive and negative control, respectively. 3. Results and discussion The AgNPs-loaded cellulose carbamate hydrogel (CCH) was denoted as AgNPs@CCH. The Tween80, CTAB, and rarasaponin modified AgNPs@CCH samples were denoted as AgNPs@CCH-T, AgNPs@CCH-C, AgNPs@CCH-R, respectively. 3.1. Carbamation of cellulose and CCH formation mechanism Hydrothermal carbamation of cellulose occurs through the substitution of functional groups; schematically, the carbamation is illustrated in Fig. 1a. Urea functional groups, specifically

9carbonyl (C=O) and amine groups (NH₂

), were substituted with primary alcohol groups (-OH, at the sixth position) of cellulose [39,47].

9In the carbamation process, the level of urea

dose affects the transparency of the hydrogel. A high dose of urea resulting in the hydrogel with high transparency, where the transparency order is mercerized filter paper (MFP) ≤ CC1 < CC3 < CC5 (most transparent). The degree of transparency of the hydrogel can be observed from the ability of the hydrogel to transmit light. Based on the results in Fig. 2b, MFP and CC1 transmissions are close to zero, indicating a lack of cellulose distribution, thus blocking light transmission. Meanwhile, CC3 and CC5 samples (with

higher carbamation levels) showed a high light transmission; this indicates proper dissolution and distribution of cellulose. High carbamation levels cause weakening of hydrogen bonds between cellulose chains so that the dissolution is better. It was found that CC5, with the ratio of cellulose to urea 1:5, has the best transparency with a degree of substitution of 0.34 ± 0.03 as calculated using the published equation [48] so that this ratio was chosen for further studies. The formation of cellulose hydrogel (without the carbamation process) has been widely reported, where the success of hydrogel formation depends on the process of cellulose dissolution [49,50]. The mechanism of the formation of CCH is similar to the formation of cellulose hydrogels, where its formation begins with the dissolution of CC in NaOH/urea solution (Fig. 1b). Next, the dissolved CC chain is cross-linked into a three-dimensional network with the help of ECH, as illustrated in Fig. 1c. The cross-linking process begins with the opening of the ECH epoxide ring through a nucleophilic attack; a basic pH condition is required in this process. Then the opened ECH ring interacts with the hydroxyl group of cellulose, which then forms interconnected networks [51–54].

3.2. Characterization of the hydrogels The effect of surfactant addition on the surface charge (without the addition of AgNPs) was investigated through its pH_{pzc} value. The plot showing pH_{pzc} is shown in Fig. S1 and the values are recorded in Fig. 5. The physical appearance of hydrogels in its swelling stage. (

1For interpretation of the references to color in this figure, the reader is referred to the web version of this article.) Table 1

. Modifications using cationic surfactant (CTAB) give high pH_{pzc} values. This is because the positively charged surfactant makes the CCH surface charge more positive so that more negative ions are needed to neutralize. In contrast, the pH_{pzc} value was lower for samples that were modified with anionic surfactants (rarasaponin). For CCH modified by a non-ionic surfactant (Tween80), the pH_{pzc} value did not show a significant difference to CCH. The FTIR spectra are presented in Fig. S2, FTIR measurements were carried out on materials without AgNPs-loaded. The peak at wave-number 1700 cm^{-1} was observed in the CCH spectra (before and after modification with surfactants); this peak does not appear in the MFP sample. This peak corresponds to the stretching vibration of the carbonyl urethane-based group (C=O), where it arises because of cellulose carbamation. The reduction in the intensity of the hydroxyl group (-OH) at wavenumber 3300 cm^{-1} was observed in CCH; this is caused by the reaction of urea with the cellulose -OH group as proposed in the mechanism of Fig. 1a [55]. Modification of CCH using different types of surfactants shows slightly different FTIR spectra. Interaction between surfactants with active groups on CCH is triggered by hydrophobic interactions that occur between hydrophobic surfactant tails and hydrophobic cellulose sites, and also hydrophilic interactions between surfactant micelles and hydrophilic cellulose groups [56,57]. In CCH-T, the peak corresponding to C]O stretch at 1718 cm^{-1} , asymmetric CH₂ stretch at 2920 cm^{-1} , and symmetrical CH₂ stretch at 2869 cm^{-1} can be observed. In CCH-C, the following peaks are found, C]O stretch at 2915 , 2857 and 1701 cm^{-1} ; tertiary amine [RN(CH₃)₃⁺] stretch at 903 cm^{-1} ; and -(CH₂)-in-plane swing at 720 cm^{-1} . In CCH-R, C]C stretch was observed at 1689 cm^{-1} . SEM images for AgNPs-loaded CCH and surfactant-modified CCH are illustrated in Fig. 3. The morphology of the cross-section of the hydrogel shows a spongiform structure with a pore diameter of around $200\text{--}500\text{ }\mu\text{m}$, and these pores facilitate space to retain water and AgNPs. The addition of surfactants and the reduction of Ag⁺ ions did not induce a significant change in the structural morphology of CCH. The presence of AgNPs on the CCH matrix was confirmed using EDX spectra (Fig. S3). All EDX spectra of the sample showed the peak of Ag; it was also seen that CCH, which was modified with surfactants, had higher Ag content. The XRD pattern for the samples is presented in Fig. 4. Diffraction peak with a high intensity, which indicates the presence of cellulose I was observed for the MFP sample. Whereas for CCH, this peak is disappeared. Instead, diffraction peaks associated with cellulose II were found in CCH samples. As stated by Aleshina et al., the dissolution of cellulose causes damage to the crystalline structure of cellulose; thus, the peak of cellulose I disappear [58]. The XRD spectra of AgNPs@CCH- R, AgNPs@CCH-T, and AgNPs@CCH-C show peaks at 2θ 38.24° and 64.44° , corresponds to (111) and (220)

); which also supported from SAED pattern, especially for AgNPs@CCH-R in inset Fig. 7d. Meanwhile, for AgNPs@CCH with no surfactant, the peak at $2\theta = 64.44^\circ$ does not present; this is due to low AgNPs content. The alteration of the XRD pattern at $2\theta = 38.24^\circ$ for both rarasaponin modified samples (AgNPs@CCH-R5 and AgNPs@CCH-R10) indicates that the presence of rarasaponin disrupts the crystalline structure of AgNPs.

3.3. Effect of surfactant on AgNPs loading

AgNPs are implanted into CCH by first soaking CCH in the AgNO₃ salt solution. Ag⁺ reduction is then initiated using NaBH₄ as a reducing agent, and thus, AgNPs are formed. The attachment of Ag⁺ ions to CCH is facilitated by the -OH group of CCH, which acts as anchor sites [59]; Fig. 6. (a) AgNPs loading on CCH, CCH-T, CCH-C, and CCH-R. Data are mean \pm SD from 2 replicates. Asterisks indicate statistical significance in comparison to CCH examined by Student's t-test; (**, P value < 0.01; *, P value < 0.05; ns, not significant)—All data show uncertainty < 5%. Elemental maps of Ag on (b) CCH and (c) surfactant-modified CCH. this interaction is reinforced by electrostatic forces [45]. Also, the cross- linked networks provide free space that allows Ag⁺ to be anchored [60]. No morphological changes were observed for Ag⁺-loaded CCH. However, visual changes can be observed, where the loading of AgNPs in CCH-C causes the color to turn purple (Fig. 5). This discoloration is caused by the formation of complexes between the Ag⁺ and the head group of CTAB [61]. Liu et al. also reported that Ag⁺ has a strong tendency to react with Br⁻ of CTAB; this is because N⁺ in the head group of CTAB cannot provide electron pairs to bind Ag⁺ [62]. The deposition of AgNPs in the hydrogel matrix can be visually confirmed by changing the color to blackish brown after reduction with NaBH₄ [63], as shown in insets Figs. 3, and 5. To measure the amount of AgNPs contained in the hydrogel, AAS measurements were carried out in addition to the EDX analysis. Before the measurement of AAS, the AgNPs-loaded hydrogels were dissolved using aqua regia. AgNPs which are released into the solution is then measured; the results of the AAS measurement are shown in Fig. 6a. It can be seen that the addition and type of surfactant result in different Ag loading capacity (LC). CCH, which is modified with the surfactant, has a higher LC value than unmodified CCH. The type of surfactant was also seen to affect LC values; modifications with Tween80 and CTAB gave lower LC values compared to rarasaponin. This is due to the anionic nature of rarasaponin, which facilitates stronger electrostatic interactions with Ag⁺. In addition, there is a probability that high surfactant concentrations (above critical micelle concentrations) allow the formation of micelles with the hydrophilic head facing out [64], and thus, create more sites to bind Ag⁺. The lower LC in CCH modified by CTAB is due to the presence of Br⁻ in solution, significantly since Br⁻ can drag out Ag⁺ ions from the hydrogel matrix. Also, cationic CTAB also causes a more favorable surface charge of CCH. Thus Ag⁺ binding was hindered. In contrast, the addition of nonionic surfactant Tween80 did not affect the CCH surface charge. Once adsorbed, the Ag⁺ ions bind with the -OH groups in the CCH matrix and surfactant micelles. These bounded Ag⁺ was reduced by the NaBH₄ that penetrates hydrogel then form AgNPs with retained interaction toward the -OH groups of CCH [65] and also with the surfactant in the modified CCH matrix. The distribution of AgNPs in the hydrogel was investigated through mapping the silver element, as shown in Fig. 6b and c. It can be seen that AgNP is well dispersed in the hydrogel matrix, and also that more AgNP is observed to be distributed in surfactant-modified CCH. The effect of surfactant on the morphology of AgNPs has been characterized. As seen in the TEM image in Fig. 7, the resultant AgNPs exhibit spherical nano-size. The histogram depicts the particle size distribution of Fig. 7. TEM images and the corresponding histograms of the particle size distribution of AgNPs on (a, a') CCH, (b, b') CCH-T, (c, c') CCH-C, (d, d') CCH-R. The inset figure shows the SAED pattern of AgNPs@CCH-R. the synthesized AgNPs@CCH with and without surfactant modification with a mean particle diameter of 11.07 ± 1.39 , 9.52 ± 0.28 , 8.87 ± 0.14 , and 8.38 ± 0.11 for CCH, CCH-T, CCH-C, and CCH-R, respectively. The AgNPs synthesized in surfactant modified CCH shows the smaller size and narrower particle size distribution than in the unmodified CCH. This may be due to aggregates remains in existence in the AgNPs prepared in the CCH without surfactant inside. A noticeable result obtained in the CCH-R that shows a significantly high relative frequency in the 5 to 10 nm range indicates a more uniform particle size.

3.4. Deformability

The mechanical properties of materials can be used as a basis for consideration in describing the strength of the

material. Deformability in units of mm/g s indicates the level of depth of the needle that squeezes CCH (mm) by the load (g) loaded per second. The higher the strength of the hydrogel, the lower the deformability value. The CCH deformability results in Fig. 8a shows that loading AgNPs causes a decrease in deformability. This indicates that the presence of AgNPs can strengthen hydrogels. Ye et al. also obtained similar results, where the compressive stress of cellulose composite hydrogel was found to increase with the addition of AgNPs [66]. A similar phenomenon was also observed in surfactant-modified CCH, where the presence of AgNPs also helped reduce the level of deformability (Fig. S4). The addition of surfactants, in contrast, exerts an effect on increasing deformability (decreased hydrogel strength). This can be attributed to the presence of surfactant micelles that act as lubricants in the hydrogel matrix. The presence of surfactant micelles in hydrogels Fig. 8. Deformability of the prepared hydrogels without (gray bar) and with AgNPs loading (black bar). Data are mean \pm SD from 3 replications. The asterisks indicate the statistical significance examined by the Student's t-test; (*, P value < 0.05; ns, not significant)—All data show uncertainty < 5%. Fig. 9. Cell viability of the prepared hydrogels with AgNPs loading for 1-day incubation. All data show uncertainty < 5%. results in reduced interaction between hydrogel molecules, and thus, reducing the strength of hydrogels [67]. The higher concentration of modifying surfactants gives a slight increase in deformability values, as shown in Fig. 8b.

3.5. Cell cytotoxicity The cytotoxicity evaluation is an extensive way of demonstrating the biocompatibility of the prepared hydrogel as a wound dressing material. Fig. 9 shows that none of the remaining cell viability was lower than 80%. According to ISO 10993-5: 1999, no cytotoxic potential in samples with cell viability higher than 70%. Therefore, it can be concluded that the AgNPs loaded hydrogels can maintain the normal function of skin fibroblasts. It can also be noted that the use of surfactant higher than CMC did not exhibit toxicity even in the high concentration of 10 mM.

3.6. Antibacterial activities The antibacterial activity of the sample was observed against gram-positive and gram-negative bacteria. The antibacterial activity is depicted from the presence of the zone of inhibition (ZOI), as shown in Fig. 10, the ZOI values are recorded in Table 2. The higher antibacterial activity is indicated by larger ZOI diameter, where the clear zones around hydrogel discs indicate that hydrogels can inhibit bacterial growth [68]. The antibacterial mechanism of AgNPs has been reported in many studies [69,70]. Briefly, Ag, which is a soft acid, tends to interact with soft bases, here, sulfur contained in protein membranes and phosphorus in bacterial DNA. Since soft acid-soft base interaction is preferable than the interaction of AgNPs with the hydroxyl groups of cellulose hydrogel, thus AgNPs will be released from the hydrogel. This released AgNP then damages bacterial cells by inducing the formation of hydroxyl radicals and penetrate into/interferes with DNA replication through the damaged cells, and thereby causing cell death. The more AgNPs come into contact, the damaging effect to the bacterial cells will be exaggerated. From the results of ZOI measurements, it was found that CCH-C had the highest antibacterial activity, followed by CCH-R > CCH-T > CCH. This is because the presence of surfactants helps increase the loading of AgNPs so that antimicrobial activity also increases. Distinct to other systems, in CCH-C—Since CTAB itself possess an antibacterial activity (as observed in negative control); thus the antibacterial activity does not solely depend on the amount of AgNPs in its matrix but also due to the presence of CTAB. The presence of surfactants also helps prevent AgNPs aggregation, thereby increasing the dispersibility of AgNPs. This increases contact between AgNPs and bacteria, thereby increasing antibacterial activity [71]. As described in most studies [72–76], hydrogels containing AgNP have a better

18inhibitory effect on the growth of E. coli

than *S. aureus*. Fig. 10. Antimicrobial activity of AgNP-loaded (a) CCH (b) CCH-T (c) CCH-C (d) CCH-R against *E. coli*; (e) CCH (f) CCH-T (g) CCH-C (h) CCH-R against *S. aureus* after incubation for 24 h at 37 °C. Scale bars are 10 mm. Table 2 Zone of inhibition (ZOI) of cellulose carbamate hydrogel (CCH) against *E. coli* and *S. aureus*. Sample Surfactant Amount of conc. (mM) antibiotic or AgNP (μ g) ZOI (mm) *E. coli* *S. aureus*

Sample	Surfactant	Amount of conc. (mM)	antibiotic or AgNP (μ g)	ZOI (mm) <i>E. coli</i>	ZOI (mm) <i>S. aureus</i>
CCH	–	–	–	0.00	61.95
CCH-T	10	5	10	15.57	71.49
CCH-C	10	5	10	14.33	75.30
CCH-R	10	5	10	14.57	75.30
Control	14.29	84.80	16.91	99.40	18.98
Control	0.00	84.33	16.45	101.50	16.87 \pm 0.76
Control	0.00	0.00	0.00	0.00 \pm 0.50	15.42 \pm 0.00
Control	0.00	0.00	0.00	12.62 \pm 0.00	0.00 \pm 0.28
Control	0.00	0.00	0.00	13.17 \pm 0.50	13.09 \pm 0.31

16.05 ± 0.38 18.73 ± 0.42 19.84 ± 0.00 0.00 ± 0.25 12.54 ± 0.42 14.37 ± 0.40 ± 0.00 ± 0.42 ± 0.00 ± 0.02 ± 0.30 ± 0.96 ± 0.42 ± 0.38 ± 0.00 ± 0.12 ± 0.35 a Amp was denoted ampicillin, the reference antibiotic used in this study. AgNPs can easily penetrate the polysaccharide layer of E. coli, plus

18the thin peptidoglycan layer of E. coli (gram-negative) causes AgNPs to

enter bacterial tissue easily [3,77]. In contrast, CCH-C showed better antimicrobial activity against S. aureus compared to E. coli. This result is in good agreement with previous studies conducted by Wieczorek et al., where CTAB showed higher antimicrobial activity against gram-positive bacteria than gram-negative bacteria [78]. Thus, CCH-C inhibition is dominated by CTAB rather than AgNPs. Comparative studies of the antimicrobial effects of AgNPs and AgNPs-hybrid materials are summarized in Table S2. As shown from the ZOI value, AgNPs@CCH-R (in this work) showed better antimicrobial activity compared to AgNPs-hybrid carboxymethyl cellulose hydrogels [79], PPEGMA-ran-PAA copolymer hydrogel [80], and bio-reduced AgNPs [81]. However, the material prepared in this work has lower activity compared to AgNPs-hybrid polycaprolactone nanofibers [82]. Nevertheless, AgNPs@CCH, which are modified by surfactants, can easily and cheaply be prepared from sustainable raw materials.

4. Conclusion Wound dressing with significant antibacterial activity and low-cytotoxicity was successfully prepared by incorporating silver nanoparticles (AgNPs) on rarasaponin-modified cellulose carbamate hydrogel (CCH-R). Incorporation of AgNPs on CCH was done by means of adsorption approach of AgNO₃ on CCH-R, followed by in situ reductions using sodium borohydride. The synergistic bacteriostatic and antibacterial effect of carbamate, rarasaponin (a natural surfactant), and AgNPs generated a hydrogel material with superior antimicrobial activity against gram-negative bacteria (E. coli) and gram-positive bacteria (S. aureus) with high biocompatibility toward skin fibroblast, L929 cell. While these findings unlocked a new approach for AgNPs loading, a limitation may arise due to the variations of the adsorption capacity of each material.

CRedit authorship contribution statement V. Bundjaja: Investigation, Data Curation, Formal Analysis, Writing – Original Draft. S. P. Santoso, A. E. Angkawijaya: Conceptualization, Investigation, Funding Acquisition, Writing – Review & Editing. M. Yuliana, F. E. Soetaredjo: Conceptualization, Resources. S. Ismadji, Y.-H. Ju: Supervision, Writing – Review & Editing. A. Ayucitra, C. Gunarto, M.-H. Ho: Data Curation, Resources.

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11Appendix A. Supplementary data Supplementary data to this article can be found online at <https://doi.org/10.1016/j.msec.2020.111542>. **References**

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8National Taiwan University of Science and Technology. Her research

inter-ests include hybrid materials for environmental and bio-technology application. Artik Elisa Angkawijaya is a Project Assistant Professor in the

1Graduate Institute of Applied Science and Technology, National Taiwan University of Science and Technology

. Her research interest span from plant metabolic adaptation to utilization of phytochemicals and under-utilized biomass for biosorbent preparation. Maria Yuliana is currently working as a lecturer and researcher

3at Department of Chemical Engineering, Widya Mandala Surabaya Catholic University

. She obtained her PhD degree (2012) from National Taiwan University of Science and Technology. Her expertise includes waste-to-energy and catalyst design. Chintya Gunarto is a Doctoral student in the

1 Department of Chemical Engineering, National Taiwan University of Science and Technology

. She has outstanding track record in research and publications. Her research works include adsorption and drug delivery system. Felycia Edi Soetaredjo is a Professor and senior researcher at the

1 Department of Chemical Engineering, Widya Mandala Surabaya Catholic University

. She obtained her PhD degree (2013) from

8 National Taiwan University of Science and Technology. Her research

interests include material designs for water and wastewater treatment. Suryadi Ismadji is a professor

3 at Department of Chemical Engineering, Widya Mandala Surabaya Catholic University

. He obtained his PhD degree (2002) from The University of Queensland, Australia. His research interests include the development of composite material for adsorption, catalyst, and drug-delivery systems. Yi-Hsu Ju, a Project Professor in the

1 Graduate Institute of Applied Science and Technology, National Taiwan University of Science and Technology

, has his research interests in the areas of Bio-chemical engineering, Bioenergy, Isolation and Identification of bio-active compounds. Ming-Hua Ho is a professor at

1 Department of Chemical Engineering, National Taiwan University of Science and Technology

. She obtained her PhD degree (2004) from National Taiwan University. Her research interests include biomaterials, biocompatibility analysis and tissue engineering. Aning Ayucitra is currently working as a lecturer and researcher

3 at Department of Chemical Engineering, Widya Mandala Surabaya Catholic University

. She obtained her PhD degree (2020) from

8National Taiwan University of Science and Technology. Her research

works include material design for adsorption and drug delivery systems. V. Bundjaja, et al.

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